A PROPOSED OECD TEST GUIDELINE FOR THE SUBMERGED MACROPHYTE, MYRIOPHYLLUM, IN A WATER-SEDIMENT SYSTEM

Jo Davies (Syngenta, UK), Margit Dollinger (Bayer CropScience AG, Germany) and Monika Ratte (ToxRat Solutions, Germany)

Introduction

Validity criteria for stats

analyses²

- Previously, the risk of herbicides to aquatic plants has been evaluated from data for four algal species and a single macrophyte species (Lemna sp.), in the US, and from data for two algal species and one macrophyte species (Lemna sp.) in the EU.
- New regulation in the EU will require data for an additional macrophyte species. Specifically, the Draft EFSA Guidance Document on tiered risk assessment for aquatic organisms indicates that data for a rooted macrophyte species may be required for substances where:
 - standard Lemna and algal EC₅₀ values are > 1 mg ai/L.
 - partitioning to sediment is a concern.
- The submerged, dicotyledonous species, Myriophyllum, has been identified as a suitable, alternative test species in light of prior experience and its known sensitivity to some herbicide chemistries.
- A protocol for testing two species of Myriophyllum in a water-sediment system was developed by an international work group and ring-tested in 15 laboratories in 2011 (Tables 1 & 2).
- In total, 51 toxicity tests were performed and the results, as well as the subsequent protocol recommendations, are summarised here.

Table 2: Test Method for Myriophyllum Species OECD draft protocol (Maltby et al, 2010) 1 Protocol Plant pots in glass test vessels (minimum volume of 2 L; Figure 1) Test system Artificial sediment (OECD 219) with added N and P nutrients Sediment Smart and Barko medium at pH 7.5 Media Via water column (method can be adapted for sediment application) **Application** Untreated control with 6 replicate test vessels and 5 concentrations, each with 3 replicate test Test design vessels; each replicate test vessel contains one plant pot of 3 shoots. 20 ± 2 °C with 16/8 hour photoperiod (160 μ E·m^{-2·}s⁻¹) Test conditions 3 to 7-day establishment (i.e. rooting) phase followed by 7-day (M. aquaticum) or 14-day (M. Test duration spicatum) exposure phase Shoot length and number of lateral branches on 3 or 4 occasions, depending on species. Fresh Biological assessments and dry weight at beginning and end of test Analytical measurements Water sampled for analysis of test substance concentration at beginning and end of test **Environmental assessments** pH, DO and temperature recorded at beginning, middle and end of test EC₅₀ and NOEC values based on yield (Y) and growth rate (Gr) derived from assessments of total shoot length (TSL), shoot dry weight (DW), shoot fresh weight (FW) and number of lateral **Endpoints** branches (LB)

¹ Maltby, L., Arnold, D., Arts, G., Davies, J., Heimbach, F., Pickl, C., Poulsen, V. (eds.), (2010). Aquatic Macrophyte Risk Assessment for Pesticides. Guidance from the AMRAP workshop in Wageningen (NL), 14-16 January 2008. ² Ratte, M. and Ratte H. T. (2012) Myriophyllum Toxicity Test, Results of a ring-test using M. aquaticum and M. spicatum grown in a water-sediment system.

CV of <35% for YFW and minimum growth rate of 0.07 d⁻¹ for FW in control plants

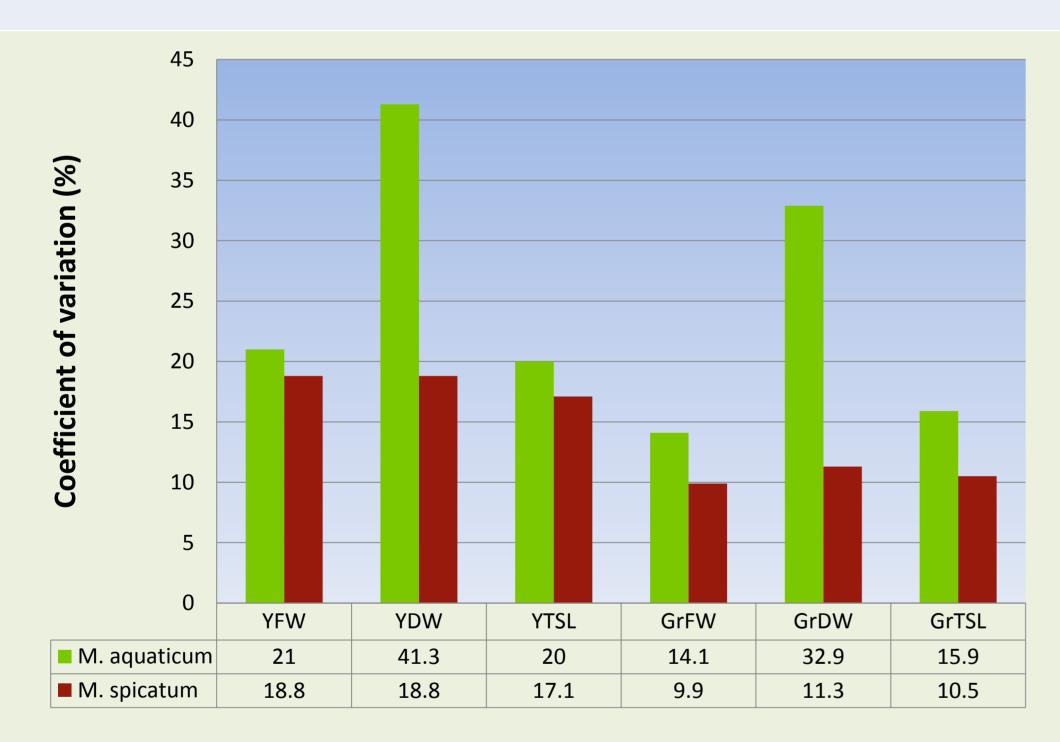


Figure 2: Mean Coefficients of Variation (%) for repeatability for control plants in valid datasets

Table 3: Mean EC ₅₀ values (mg/L) for valid datasets							
Test substance	Assessment parameter	M. aquaticum			M. spicatum		
		7-d EC ₅₀	n	95% PI	14-d EC ₅₀	n	95% PI
3,5-DCP	YDW	-	0	-	5.0	5	2.3 – 10.7
	GrDW	3.7	1	n.a.	4.8	4	2.4 – 9.7
	YFW	4.6	2	3.6 – 5.8	4.7	6	2.0 – 10.7
	GrFW	5.2	2	2.8 – 9.7	5.3	5	2.5 – 11.2
	YTSL	5.7	5	3.1 – 10.6	5.3	5	2.9 – 9.8
	GrTSL	6.1	5	3.5 – 10.8	6.3	6	3.9 – 10.1
Isoproturon (IP)	YDW	0.12	1	n.a.	0.052	4	0.016 - 0.168
	GrDW	0.13	1	n.a.	0.074	4	0.034 - 0.163
	YFW	0.86	2	0.59 – 1.25	0.083	6	0.024 - 0.280
	GrFW	1.18	2	1.09 – 1.27	0.164	6	0.046 - 0.583
	YTSL	0.37	3	0.16 – 0.84	0.140	6	0.054 - 0.362
	GrTSL	0.51	3	0.13 - 1.91	0.315	6	0.117 - 0.849
Trifluralin (TF)	YFW	-	-	-	0.319	2	0.104 - 0.978
	GrFW	-	-	-	0.840	1	n.a.
	YTSL	0.82	3	0.16 – 4.1	0.165	3	0.115 - 0.23
	GrTSL	0.75	1	n.a.	0.314	3	0.203 - 0.48

n.a. not applicable - not determined due to lack of valid datasets (3,5 DCP) or the absence of a consistent dose response (TF)

Table 1: Ring-test Details

Test species M. aquaticum

Test substances

Isoproturon (IP)

Trifluralin (TF)



3,5, dichlorophenol (3,5 DCP)



M. spicatum

Alterra, Wageningen, NL

BASF SE, Limburgerhof, DE

Participating laboratories

- Bayer CropScience LP, Stilwell, USA
- Biochem Agrar GmbH, Gerichshain, DE
- Chemex Environmental International Ltd, Cambridge, UK
- Dr U Noack Laboratorien, Sarstedt, DE
- ECT Ökotoxikologie GmbH, Flörsheim, DE
- Eurofins Agroscience Services GmbH, Niefern-Öschelbronn, DE
- Federal Environmental Agency, Berlin, DE
- Fraunhofer IME, Schmallenberg, DE Harlan Laboratories Ltd, Itingen, CH
- IBacon GmbH, Rossdorf, DE Institute of Industrial Organic Chemistry, Pszczyna, PL
- University of Novi Sad, SRB
- Smithers Viscient, Wareham, USA



Figure 1: Myriophyllum spicatum test at US Army Corps of Engineers

Key Findings

Performance of control plants and reproducibility (Figure 2)

- Analyses of all datasets indicated that 86% of *M. spicatum* tests and 50% of *M. aquaticum* tests met the preliminary validity criteria for analysis.
- The valid datasets showed control CVs of up to ca. 20%, although *M. aquaticum* CVs were >30% for DW variables. The great variability seen for *M. aquaticum* was attributed to poor growth in some tests.
- Lateral branches were formed in *M. spicatum* but infrequently in *M. aquaticum*, prohibiting the derivation of endpoints based on this parameter for *M. aquaticum*.
- Generally, both species allowed statistical testing at minimal detectable differences of up to 30%, with the exception of the DW variable for *M. aquaticum*.

Sensitivity to 3,5-DCP, Isoproturon and Trifluralin (Table 3)

- Both species were sensitive to all test substances although DW in both species and FW in M. aquaticum did not show a conclusive response to TF. This lack of consistent sensitivity may be attributed to the poor water solubility of TF (0.22 mg/L).
- Both species showed similar sensitivity to 3,5-DCP while *M. spicatum* was more sensitive than *M.*
- aquaticum to IP and TF. Systematic differences between Gr and Y endpoints based on DW, FW or TSL were not apparent, except for
- Growth rate and Y based on lateral branch formation (M. spicatum) were not typically the most sensitive endpoints for any test substance.
- Root development was affected by all test substances in both species. Assessments of M. aquaticum root length indicated that root endpoints showed similar sensitivity to shoot endpoints for 3,5-DCP (n=2) but were less sensitive than shoot endpoints for IP (n=1).

Recommendations included in the Updated Draft OECD TG

the finding that Gr endpoints tended to be higher than Y endpoints.

Steps to improve test performance, reproducibility and efficiency

- Use uniform, healthy plant material and verify root formation prior to test initiation.
- Extend the exposure phase for *M. aquaticum* to 10 days if required to achieve doubling.
- Use submerged M. aquaticum shoot material, if possible, to initiate test cultures.
- Reduce the number of assessments during the test period, i.e. 3 measurements of TSL for both species.

Validity criteria

- Total shoot length and shoot fresh weight in control plants must at least double within the test duration.
- The mean coefficient of variation for Yield, based on measurements of shoot FW, DW and length, in the control cultures must not exceed 35%.

Recommended Endpoints

- The primary recommended endpoints will be based on growth rate. Options are included for the estimation of yield-based endpoints.
- Growth rate and yield endpoints will be derived from TSL, DW and FW.

Status of Draft OECD TG for Myriophyllum in a water-sediment system

- The Draft TG has been updated to include these recommendations and has been circulated to all participants and interested parties for comment.
- The updated TG will be made available to OECD during May 2013.

Acknowledgements

This work is co-ordinated by the AMRAP *Myriophyllum* Workgroup:

Jo Davies (Co-chair; Syngenta), Margit Dollinger (Co-chair; Bayer CropScience), Gertie Arts (Alterra), Eric Bruns (Bayer CropScience), Nina Cedergreen (Copenhagen University), Peter Dohmen (BASF), Ute Feiler (Federal Institute of Hydrology, DE), Mark Hanson (Manitoba), Udo Hommen (Fraunhofer, IME), Katja Knauer (BLW, CH), Johanna Kubitza (BASF), Dirk Maletzki (UBA), Lorraine Maltby (Sheffield University), Angela Poovey (US Army Corps of Engineers).